

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of

GUERIN-MARCHAND ET AL

Serial No.:09/837,344

Filed: April 19, 2001

For: PEPTIDE SEQUENCES
SPECIFIC FOR THE HEPATIC
STAGES OF P. FALCIPARUM
BEARING EPITOPES CAPABLE
OF STIMULATING THE T
LYMPHOCYTES

Group Art Unit: 1645

Examiner: N. Minnifield

For: PEPTIDE SEQUENCES

SPECIFIC FOR THE HEPATIC

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1.132 DECLARATION

Hon. Commissioner of Patents P.O. Box 1450 Alexandria, Virginia 22313-1450

I, Pierre Druilhe do hereby declare the following:

- (1) I am currently the director of the Biomedical Parasitology Unit at Institut Pasteur in Paris, France and have been the director of this department for many years. As can be seen from my attached Curriculum Vitae, I have published 260 articles and have 10 U.S. Patents issued in my name. I am one of the inventors of the abovecaptioned U.S. patent application.
- (2) I have read the last U.S. Official Action dated February 10, 2005, for the above-identified patent application. It is my understanding that the Examiner deems that the claims directed to vaccines cannot be produced by a scientist using the disclosure of the above-captioned specification since the specification does not teach a scientist how to obtain a malaria vaccine. It appears that the Examiner's reasoning in maintaining this rejection is that the specification purportedly only describes the construction of a Genomic DNA Library and the

- provides further immunological testing of the chosen clones, as well as conclusions reached by this testing. I respectfully disagree with the Examiner for the following reasons.
- (3) The specification clearly describes immunological responses of subjects exposed to malaria. More specifically, the immunization of mice with the LSA-R-NR proteins and LSA-R peptides of mice of different haplotypes with peptides LSA-R, LSA-J and LSA-NR and responses of the lymphocytes of subjects exposed to malaria indicated that a T-epitope existed in the LSA-R peptide, as well as a B epitope. Other results with more than 500 individuals exposed to malaria should that this T epitope was recognized by the antibodies of about 95% of the subjects studied in Senegal, Leppervoln, Madagascar and Kenya. Responses of lymphocytes obtained from 5 adult African subjects exposed to malaria and more than 200 adult African subjects confirmed that a T epitope of the LSA molecule was defined by the amino acid contained in the synthetic sequence of the peptide LSA-NR.
- (4) Lymphocytes of chimpanzees, which were immunized using the LSA-R-NR recombinant protein (SEQ ID NO:19), confirmed that this protein contained a T epitope. In this regard, the chimpanzees were immunized at 15 day intervals and production of antibodies specific for LSA-R-NR was demonstrated. 60% of the lymphocytes were of the CD8+ phenotype, which corresponds to cytotoxic T lymphocytes.
- (5) The specification also discloses studies on mice that were injected with the recombinant protein LSA-R-NR and proof of B epitopes in the LSA-R-NR protein. Further studies conducted in sera taken from 120 African subjects confirmed that the peptide LSA-NR has a B epitope and that this epitope was recognized by 65% of the subjects.
- (6) The specification also discloses that chimpanzees were immunized with LSA-R-NR and then further injected with an intravenous injection of 28 million sporozoites of *P. falciparum*. Control chimpanzees in which no immunization occurred were also inoculated in the same manner. Liver biopsies of the immunized chimpanzees were taken on the 6th day after infection. The biopsies of the immunized chimpanzees showed the existence of cellular reaction, lympho-monocytic, around

- the hepatic schizonts, infiltrating the schizonts and capable of destroying them. Such images were not observed in the control group.
- (7) The cytolytic capacity of the lymphocytes on the immunized chimpanzees in paragraph (6) was further analyzed using a chromium 51 label. The results illustrated that the T epitopes are capable of activating cytolytic T lymphocytes for the LSA-R-NR sequence.
- (8) Thus, for the results obtained in the above paragraphs (3) to (7) the following was concluded about the LSA-R-Nr (536) polypeptide:
 - (a) that this polypeptide is recognized by antibodies originating from subjects with malaria;
 - (b) that this polypeptide is recognized by sera of chimpanzees immunized therewith;
 - (c) that this polypeptide induces the function of antibodies (B epitopes);
 - (d) that this polypeptide induces proliferative responses of T lymphocytes indicating the presence of T epitopes in both chimpanzees and man;
 - (e) that the polypeptides NR and TER include a major T epitope and a B epitope for man as well as for mouse and the chimpanzee.
- (9) Further studies were undertaken with Aotus monkeys confirming the presence of these B and T epitopes and the good antigenicity, as demonstrated in Perlaza et al, Infection and Immunity July 1998 p. 3423-3428, previously of record. In fact this paper indicates that in various types of monkeys such as Aotus, Saimiri and Cebus monkeys liver schizogony can be obtained with P. falciparum strains without the need for previous adaptation to these monkeys. Thus, this animal model can be used for preclinical development of pre-erythrocytic malaria vaccines.
- (10) Therefore, since the peptide LSA-R-NR (SEQ ID NO:19) contains both T and B epitopes, larger sequences such as those of SEQ ID Nos 39-42 and 43-46, which contain the LSA-R-NR sequence also contain both T and B epitopes.

- (11) Thus, the present specification clearly demonstrates that several polypeptides obtained by the methods in the specification have T epitopes, B epitopes or both and thus induce a wide range of immune responses.
- (12) It appears that the Examiner has taken a strict interpretation of the word "vaccine" since at page 6 of the Official Action, the Examiner states that "the definition of a vaccine is a product that provides protection (prophylaxis) against infection, in this case protection against malaria." However, it is well known that a vaccine can either block the effects of the pathogen toxins or prime the immune system against the pathogen such that infection is brought under control more quickly. The latter is done through the vaccine's capability to stimulate antibody (B cells) and T cell responses that can then respond quickly to infection and prevent the pathogen from causing clinical illness.
- (13) Indeed, vaccination works by producing T cell and B cell responses. Thus, when vaccinated cytotoxic T cells are produced by the immune system which recognizes the diseased cells and helper T cells assist in activating killer T cells and stimulate B cells, thus producing a cell-mediated immune response. The B cells secrete antibodies which attack malaria, thus producing a humoral immune response.
- (14) Thus, it can be concluded that by discovering polypeptides having T epitopes and B epitopes as set forth in the present specification, a skilled artisan can produce a vaccine composition based on the disclosure of the specification. Indeed, I come to this conclusion, since additional studies have been made with LSA-1 antigens in endemic areas since the filing of the present patent application which have linked the LSA-1 antigen with protective immunity. More particularly B-cell and T-cell epitopes in LSA-1 have been associated with protective immunity as stated in the attached Document 1.

I also declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

28 Narch earl

Date

Plerre DRUILHE



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1975 - Post-graduate degree in Immunology (University of Paris)

ACADEMIC APPOINTMENTS:

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1976 - Chef de Travaux Pratiques

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1985 - Praticien hospitalier of Paris Hospitals

1975 - Assistant of Paris Hospitals

1972 - Resident of Paris Hospitals

C - Research Institute

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HIV/AIDS

Disease burden. WHO and UNAIDS have estimated that at the end of 2002 the number of adults and children living with HIV/AIDS worldwide will have reached 42 million (AIDS Epidemic Update 2002). It is also estimated that, during 2002, 5 million people (including 800 000 children aged under 15 years) became infected and 3.1 million have died of the disease (WHO, 2002). Human Immunodeficiency viruses belong to the Retroviridae family, lentivirus genus. Two types have been desbribed: HIV-1 and HIV-2. HIV infections are now almost equally distributed between men and women, with an estimated 17.6 million women aged 15-49 living with HIV/AIDS. The vast majority of people living with HIV/AIDS are not aware that they are carrying the virus. Deaths in women also continue to increase, accounting for an estimated 48% of adult deaths due to HIV in 2002. HIV/AIDS is the leading cause of death in sub-Saharan Africa and the fourth biggest killer worldwide.

Some regional trends can be observed. During 2002, AIDS killed 2.4 million people In sub-Saharan Africa while an estimated 3.5 million people have been newly infected with HIV, bringing the number of Africans living with HIV/AIDS to 29.4 million. sub-Saharan Africa remains the hardest-hit region, accounting for 68% of the 5 million people infected worldwide with HIV during 2001, 71% of the people living with HIV/AIDS and 77% of AIDS deaths.

The estimated number of adults and children living with HIV in Latin America and the Caribbean at the end of 2002 was 1.5 million. While in some countries HIV infections remain concentrated mainly in men who have unprotected sex with other men and injecting drug users, others are experiencing increasing rates of heterosexual transmission. AIDS mortality has been reduced in some countries thanks to antiretroviral therapy.

Eastern Europe and Central Asia continue to experience the world's fastest-growing HIV/AIDS epidemic. During 2002, there were an estimated 250 000 new infections, bringing the number of people living with HIV/AIDS to 1.2 million. Most of the infections continue to occur among injecting drug

In North Africa and the Middle East, the number of people living with HIV/AIDS now totals 550 000. While HIV prevalence continues to be low in most countries in the region, an increasing number of HIV infections have been detected in several countries, particularly in countries experiencing complex emergencies.

During 2002, highly active antiretroviral therapy has continued to reduce progression to AIDS, deaths and HIV transmission from mother to child in the Industrialized countries of North America, Western Europe and the Pacific.

However, these successes in treatment and care are not being matched by progress in prevention. During 2002, 76 000 individuals became infected with HIV in industrialized countries, where an estimated 1.6 million people are now living with HIV. New evidence of rising HIV infection rates is emerging, particularly in marginalized communities.

Vaccines. While HIV/AIDS continues to spread in all regions of the world, there are positive signs. In both industrialized and developing countries, an increasing number of HIV-positive people can live longer and healthier lives thanks to antiretroviral theraples. Large-scale prevention programmes have reversed epidemic trends in some Asian countries. The estimated number of new infections in most Central/East/West African countries seem to decline, and there are some initial indications that the epidemics might have peaked In southern Africa. But above all, a new determination to fight the epidemic has emerged following the United Nations General Assembly Special Session on HIV/AIDS in July 2001. However, despite these encouraging trends, a preventive vaccine is more than ever needed, in particular for developing countries. The development of a safe and effective vaccine is hampered by the high genetic variability of HIV, the paucity of knowledge on the immune mechanisms of protection, the absence of relevant and predictive animal models, and the complexity of the implementation of efficacy trials, especially in developing countries . Several vaccine candidates have been tested over the past 15 years and are in the pipeline for further human testing. The first Phase I trial of an HIV vaccine

Diarrhoeal diseases Rotavirus Enterotoxigenic Escherichia <u>coli</u> Shigella Cholera Typhoid Calciviruses

Acute respiratory infections Streptococcus pneumoniae Respiratory syncytial virus Parainfluenza virus type 3 Influenza

Tuberculosis Buruli Ulcer Chlamydia Trachomatis HIV/AIDS Herpes simplex virus type 2 Malaria Leishmaniasis Schistosomiasis Dengue Japanese Encephalitis Tick-Borne Encephalitis **West Nile Encephalitis** Meningitis N. meningitidis Hepatitis C Hepatitis E Helicobacter pylori Human Papilloma Virus Epstein Barr Virus Poliomyelitis Measies Rabies Group A Streptococcus Group B Streptococcus Leptospirosis

R&D status for new

vaccines and biologicals

was conducted in the US in 1987. Since then, over 30 candidate vaccines have been tested in over 80 Phase I/II clinical trials, involving over 10 000 healthy human volunteers (adults and infants). The majority of these trials have been conducted in the US and Europe, however, trials have also been conducted in developing countries (Brazil, China, Cuba, Haiti, Kenya, Thailand, Trinidad and Uganda). The effort to develop and evaluate HIV vaccines must increase, especially in Africa. This effort will be strengthened by the African Aids Vaccine Programme (AAVP), which has been recently established following an initiative in WHO and UNAIDS. Only two efficacy trials have been started so far, both using the same approach of a monomeric gp120, one in the United-States (with sites in Canada and in the Netherlands), the other in Thailand (Vaxgen/CDC). Definite results from the USA trial have been reported in March 2003. The study did not show a statistically significant reduction of HIV infection within the study population as a whole, which was the primary endpoint of the trial. However, the study did show a statistically significant reduction of HIV infection in certain vaccinated groups. Protection appeared to correlate with the higher level of vaccine-induced neutralizing antibodies observed in these groups, according to Vaxgen. A third efficacy trial of a recombinant canarypox HIV vector prime-gp120 boost vaccine in heterosexual volunteers in Thailand is expected to start late 2003 (WRAIR/NIH/Vaxgen/Aventis Pasteur). Other interesting approaches based on DNA prime and recombinant poxviruses (MVA, fowlpox) boost are being tested in humans. Recombinant adenoviruses represent another promising approach, already tested in Phase I in humans. Other candidate vaccines include recombinant Salmonella (IAVI/Institute for Human Virology, University of Maryland, USA), VEE (Alphavax, USA), subunit HIV proteins (GSK, Center for Genetic Engineering and Biotechnology (CIGB) in Cuba, among others), DNA vaccines (Wyeth/University of Pennsylvania, among others) or peptides (Wyeth/Duke University). The development of a safe, effective, and affordable HIV vaccine remains the scientific and public health challenge of this new century. However, it is urgent that the availability and affordability of a safe and efficient HIV vaccine should by any means strongly encourage, strengthen, and expand the efforts of traditional prevention against HIV infection proven to be efficient in some countries.

WHO-UNAIDS HIV Vaccine Initiative Back to top

Herpes simplex virus type 2

Disease burden. HSV-2 prevalence is increasing worldwide. There is now ample evidence that Herpes simplex virus type 2 (HSV-2) infection, the most common cause of genital ulcers worldwide, is a major cofactor favoring HIV infection. HSV-2 prevalence is generally higher in developing than in developed countries and in urban than rural areas. The HSV-2 prevalence in developing countries, although very high, varies widely according to the countries, the gender, urban versus rural areas, ranging from 2-74%, very few being available from Asian and South American countries. HSV-2 incidence data are scarce. In the community-based survey, HSV-2 prevalence increased with age until 25 years, leveled off at 50% in both genders. The same independent predictors of HSV-2 infection were identified in both genders: older age, higher lifetime number of sexual partners, positive HIV serology, and positive Treponema pallidum hemagglutination serology. In Bangladesh a study conducted in truck drivers showed a high HSV-2 prevalence of disease were HSV-2 (25.8%), compared to serological syphilis (5.7%), gonorrhea (2.1%), chlamydia (0.8%). In New Zealand, increased rates of HSV-2 acquisition after age 21 may be due to a higher prevalence of infection in the pool of potential partners encountered during the third decade of life. Factors related to partner choice may have more influence on the risk of HSV-2 infection than the number of sexual partners alone. Overall prevalence is higher in women compared with men, especially among the young , and rates up to 40 % have been reported among women aged 15-19 in Kisumu (Kenya). Prevalence is higher in the USA (22% in adults) compared to Europe (generally lower than 15%). It is now estimated that in the USA alone, 40 to 60 million people are HSV-2-infected, with 2 million incident cases per year and 600 000 clinical cases. The clinical spectrum of HSV-2 includes primary infection, a first episode of genital herpes and recurrent episodes of clinical disease (4-5 per year). In addition, subclinical infection may be associated with infectious viral shedding. The proportion of infections which are sympomatic is estimated to be between 13 and 37%, although this is higher in HIV positive individuals. The risk of neonatal herpes is very low (less than 3%) among HIV-negative pregnant women living in developed countries, but few data are available on neonatal herpes in developing countries. A recent meta-analysis of thirty-one studies addressing the risk of HIV infection in HSV-2-seropositive persons was performed. For nine cohort and nested case-control studies that documented HSV-2 infection before HIV acquisition, the risk estimate was 2.1 (95% confidence interval, 1.4-3.2). Thus, the attributable risk percentage of HIV to HSV-2 was 52%, and the population attributable risk percentage was 19% in populations with 22% HSV-2 prevalence but increased to 47% in populations with 80% HSV-2 prevalence. For 22 case-control and cross-sectional studies, the risk estimate was 3.9 (95% confidence interval, 3.1-5.1), but the temporal sequence of the two infections cannot be documented. Control strategies for HSV-2 need to be incorporated into control of sexually transmitted infections as a strategy for HIV prevention.

In developed countries, acquisition of HSV-1 in childhood has decreased as HSV-2 seroprevalence has increased, suggesting a possible protective effect of HSV-1 against HSV-2 acquisition. However, studies have shown discrepant results in this respect. Although HSV-1 does not seem to modify the risk of HSV-2 acquisition, It seems to increase the proportion of asymptomatic seroconversions and, in one study, to increase the rate of HSV-2 shedding. Infection with HSV-1 in childhood is almost universal in developing countries, where HSV-2 prevalance is very high, and this confirms that HSV-1 provides limited protection against HSV-2 infection.

Vaccines. Herpesvirus 2 belongs to the enveloped *Herpesviridae* family. The first generation of vaccines was recombinant subunit viral glycoproteins. <u>Chiron</u> developed a two-component (30 mg of both gB2 and gD2 glycoproteins) subunit vaccine formulated in MF59 adjuvant and SmithKline Beecham a monovalent vaccine (gD2) formulated in alum + monophosphoryl lipid A (Corixa). The <u>Chiron vaccine</u> induced very high antibody titres, and efficacy in women for the first 5 months was 26%, but this protection against

infection was not sustained. Chiron doesn't have an active HSV-2 vaccine at present. The Phase III trials of the vaccine developed by GlaxoSmithKline (glycoprotein gD2 formulated with adjuvant) showed limited efficacy, depending on gender and previous exposure to HSV-1. Indeed, these trials showed a 73% and 74% efficacy (P=0.01 and 0.02, respectively) against genital herpes disease in HSV-1- and HSV-2-negative women. Trends towards protection in women against HSV infection were also seen in both studies (39-48% efficacy), although not statistically significant. The main disadvantages of this vaccine are the apparent failure to improve on protection provided by HSV-1 infection and the need for frequent administration to boost host immunity. Further efficacy trials of this vaccine, which has already been administrated In about 7 500 individuals, are pending in collaboration with NIAID. A novel HSV-2 candidate vaccine has been developed by Cantab Pharmaceuticals (now Xenova)/GlaxoSmithKline based on a genetically Disabled Infectious Single Cycle (DISC, glycoprotein H-deleted, ICP8 gene mutation) replicative vaccine, which is believed could have higher efficacy than previous vaccines. This new candidate vaccine has been tested in Phase II trials in the US and UK, showing good tolerance and inducing neutralizing antibodies and CTL in 83% of the vaccine recipients. Nevertheless, no difference in time to recurrence was observed in this therapeutic candidate vaccine in HSV-2 seropositive symptomatic, and no difference was recorded in shedding. Therefore, Xenova (without GSK) is refocusing their programme on prophylactic applications for their DISC vaccine. One complexity of evaluating protection against infection induced by the DISC vaccine is that, because of the similarity of the disabled virus and wild HSV-2, it will not be possible to distinguish natural infection from vaccine-induced Immunity. Another live, replication-impaired vaccine is currently under development by Avant Immunotherapeutics. AuRx, Inc concentrate on live genetically-attenuated replication-competent vaccines and PowderJect and Merck on DNA vaccine formulations.

A vaccine which protects only women would be expected to reduce HSV infection and disease in vaccinated women, decrease the rate of neonatal HSV infection, have an impact on the epidemic spread of genital herpes in men and women, and finally possibly reduce acquisition and transmission of HIV infection. Failure to protect HSV-1 seropositive women may result if vaccination does not add to the natural protection provided by HSV-1. In this case administration of vaccine to young children, before HSV1 occurs, would not be particularly helpful. Lack of efficacy of vaccines in HSV1-infected individuals would render the vaccine useless in developing countries, where HSV-1 infection is almost universal.

IVR Herpes simplex virus type 2 section Back to top

Malaria

Disease burden. Malaria is by far the world's most important tropical parasitic disease, and kills more people than any other communicable disease except tuberculosis. Malaria is a public health problem today in more than 90 countries, inhabited by a total of some 2 400 million people - 40% of the world's population. Malaria is endemic in a total of 101 countries and territories: 45 countries in WHO's African Region, 21 in Its Americas Region, 4 in its European region, 14 in its Eastern Mediterranean Region, 8 in its South-East Asia Region, and 9 in WHO's Western Pacific Region. Worldwide prevalence of the disease is in the order of 300-500 million clinical cases each year. More than 90% of all malaria cases are in sub-Saharan Africa.

Mortality due to malaria is estimated to be over 1.1 million deaths each year (WHO, 2002). The vast majority of deaths occur among young children in Africa, especially in remote rural areas with poor access to health services. Malarla kills one child every 30 seconds. This preventable disease has reached epidemic proportions in many regions of the world, and continues to spread unchecked. In absolute numbers, malaria kills 3 000 children under five years of age per day. It is a death toll that far exceeds the mortality rate from AIDS. African children under five years of age are chronic victims of malaria, suffering an average of six bouts a year. Fatally afflicted children often die less than 72 hours after developing symptoms. In those children who survive, malaria also drains vital nutrients, impairing their physical and intellectual development. Malarial sickness is also one of the principal reasons for poor school attendance. Other high-risk groups are women during pregnancy, and non-immune travelers, refugees, displaced persons and laborers entering endemic areas. During pregnancy, malaria causes severe anemia, and is a major factor contributing to maternal deaths in malaria endemic regions. Pregnant mothers who have malaria and are HIV-positive are more likely to transmit HIV to their newborn. In many developing countries and in Africa especially, malaria exacts an enormous toll in lives, in medical costs, and in days of labor lost. The causative agents in humans are four species of Plasmodium protozoa (single-celled parasites) - P.falciparum, P.vivax, P.ovale and P.malariae transmitted by Anopheline mosquitoes, the number and type P.falciparum, P.vivax, P.ovale and P.malariae transmitted by Anopheline mosquitoes, the number and type of which determine the extent of transmission in a given area. Transmission of malaria is affected by climate and geography, and often coincides with the rainy season. Of these, P.falciparum accounts for the majority of infections and is the most lethal. Malaria is a curable disease if promptly diagnosed and adequately treated.

The geographical area affected by malaria has shrunk considerably over the past 50 years, but control is becoming more difficult and gains are being eroded. Malaria's reach is spreading. In malaria endemic parts of the world, a change in risk of malaria can be the unintended result of economic activity or agricultural policy that changes the use of land (e.g. creation of dams, irrigation schemes, commercial tree cropping and deforestation). "Global warming" and other climatic events such as "El Niño" also play their role in increasing risk of disease. The disease has now spread to highland areas of Africa, for example, while El Niño events have an impact on malaria because the associated weather disturbances influence vector breeding sites, and hence transmission of the disease. Many areas have experienced dramatic increases in the incidence of malaria during extreme weather events correlated to El Niño. Moreover, outbreaks may not only be larger, but more severe, as populations affected may not have high levels of immunity. Quantitative leaps in malaria incidence coincident with ENSO (El Niño/Southern Oscillation) events have been recorded around the world: in Bolivia, Columbia, Ecuador, Peru and

Venezuela in South America, in Rwanda in Africa, and in Pakistan and Sri Lanka in Asia.

More than any other disease, malaria hits the poor. Malaria endemic countries are some of the world's poorest. Costs to countries include costs for control and lost workdays - estimated to be 1-5% of GPD in Africa. For the individual, costs include the price of treatment and prevention, and lost income. Rural communities are particularly affected. In rural areas, the rainy season is often a time of intense agricultural activity, when poor families earn most of their annual income. Malaria can make these families even poorer. In children, malaria leads to chronic school absenteeism and there can be impairment of learning ability. Urban malaria is increasing due to unplanned development around large cities, particularly in Africa and South Asia.

The estimated costs of malaria, in terms of strains on the health systems and economic activity lost, are enormous. In affected countries, as many as 3 in 10 hospital beds are occupied by victims of malaria. In Africa, where malaria reaches a peak at harvest time and hits young adults especially hard, a single bout of the disease costs an estimated equivalent of 10 working days. Research indicates that affected familles clear only 40% of land for crops compared with healthy families. Knowledge about malaria is markedly low among affected populations. In one recent survey in Ghana, for example, half the respondents did not know that mosquitoes transmit malaria. The direct and indirect costs of malaria in sub-Saharan Africa exceed US\$ 2 billion, according to 1997 estimates. According to UNICEF, the average cost for each nation in Africa to implement malaria control programmes is estimated to be at least US\$ 300 000 a year. This amounts to about six US cents (US\$ 0.06) per person for a country of 5 million people.

A limited number of drugs for treatment of malaria are available today. Due to worsening problems of drug resistance in many parts of the world, adequate treatment of malaria is becoming increasingly difficult. Although some new drugs have appeared in the last 20 years (e.g., mefloquine, halofantrine, artemisinin derivatives, malarone, atovaquone + proguanil, co-artemether), new (especially inexpensive and affordable) drugs and more practical formulations of existing drugs/compounds are badly needed.

Vaccines. International efforts to combat malaria are unprecedented (see WHO website) through a Global Malaria Control Strategy whose activities are coordinated by WHO's Programme on Communicable Diseases (CDS), Roll Back Malaria coordinating the efforts of four UN-System agencies (UNDP, UNICEF, WHO and the World Bank) launched on 30 October 1998, Multilateral Initiative on Malaria (MIM) launched in Dakar in January 1997 when a number of institutions (from both public and private sectors) joined forces to promote malaria research in Africa. The UNDP/World Bank/WHO Special Programme on Tropical Diseases (WHO/TDR) has joined the Initiative, establishing a Task Force to address the needs of endemic countries and to fund activities related to strengthening research capacities in malaria. The Task Force has mobilized around 40 countries and 161 partners for submitting proposals for review. Fifteen partnership projects involving 20 African and 5 European countries and the USA have been funded. The main malaria vaccine funding agencies are the USA <u>NIH</u>, the <u>European Union</u>, either directly of through the <u>European Malaria Vaccine Initiative (EMVI)</u>, the <u>USAID</u>, the <u>Malaria Vaccine Initiative (MVI)</u> and Rockefeller Foundation . There are four general categories of malaria vaccine candidates, each representing a different stage of intervention. Virtually all the malaria vaccine candidates (with the exception of GPI anchor antigen described below) are cell surface antigens present during one of the three developmental stages of the Plasmodium parasite. Pre-erythrocytic (sporozoite) vaccines are those directed against the sporozoite and liver stages of the malaria parasite. The sporozoite is the form of the parasite introduced into the human host by the bite of an infected mosquito and that invades liver cells. A sporozoite vaccine could prevent infection either by blocking invasion of liver cells (antibody response) or destroying infected liver cells (cell-mediated response) by preventing release of parasites into the bloodstream. The asexual blood-stage (erythrocytic) vaccines are directed against the merozoite stage of the parasite, which invades and replicates in the red blood cells. A blood-stage vaccine would be expected to reduce both the severity and duration of the disease by decreasing the blood parasite density, which correlates with reduced disease symptoms and risk of death. The transmission-blocking vaccines are designed to raise antibodies (in humans) against the gamete stage of the parasite present in the mosquito gut. Such antibodies taken up by a mosquito during a blood meal should block further parasite development in the mosquito, becoming a non-infectious vector. Blocking transmission of the parasite could reduce infectivity of the mosquitoes (carrying fewer parasites) and extend the useful life of a pre-erythrocytic or blood-stage vaccine by preventing transmission of antibody-resistant mutants. A fourth type of potential malaria vaccine is an anti-disease vaccine. This approach to a vaccine involves the identification of parasite toxins that contribute to disease. The glycosylphosphatidyl inositol (GPI) anchor, which tethers several of the parasite antigens to the membrane, has been shown to be highly toxic in mouse models. GPI as a vaccine would have to be detoxified enough to be safe but the potential for disease attenuation with this approach is real. Most investigators now acknowledge that a combination vaccine (multi-antigen, multi-stage) will probably be the best approach to effective vaccination. The various malaria vaccine candidate antigens are expressed and manufactured in a number of different ways including recombinant protein, DNA vaccine constructs, viral-vectored constructs, synthetic peptides, and chimeric proteins. The Plasmodium genome is very A-T rich, unlike most of the microbial organisms (bacteria, yeast or virus) or animals used to express recombinant parasite antigens, and the organism has quite different codon usage. Enhanced expression of recombinant Plasmodium antigens may be obtained by creating synthetic genes using optimized codons according to the organism used for expression.

The most advanced and well-documented pre-erythrocytic (liver-stage) vaccine candidates are derived from the circumsporozoite (CS) antigen present on the sporozoite. Such a vaccine candidate developed by <u>GlaxoSmithKline</u> and the <u>Walter Reed Army Institute of Research (WRAIR)</u>, is referred to as RTS,S/ASO2. This vaccine is comprised of the antigenic C-terminus (amino acids 207-395) of the CS gene from P. falciparum fused to the hepatitis B surface antigen. This chimeric polypeptide containing the hepatitis B surface antigen is co-expressed with hepatitis B surface antigen in Saccaromyces cerevisiae. Initial Phase I clinical trials of RTS,S formulated with the ASO2 adjuvant (containing MPL, QS21 and an

oil-in-water emulsion) showed protection against sporozoite challenge in 6 out of 7 volunteers. More recently, a dose-range Phase I/II study showed levels of efficacy from 30% (single dose) to 55% (3 doses). Overall protective efficacy was 41% among 41 vaccinees. Further trials in a pediatric population in The Gambia are now in progress (www.malariavaccine.org). Current studies are almed at combining RTS,S with the blood-stage antigen MSP1 (see below).

Another CS-based vaccine candidate includes a 102-amino acid synthetic peptide representing the antigenic C-terminus of the circumsporozoite antigen. The peptide was safe and immunogenic in humans (no challenge). This vaccine candidate will soon enter Phase I/II trials for safety and efficacy in Europe. In addition, Apovia (USA) is developing with MVI funding a vaccine based on CS determinants expressed on Hepatitis B core particles This vaccine should soon enter clinical development.

The Department of Defense (USA) in collaboration with <u>Vical, Inc</u>. is developing DNA vaccines for malaria (the Multi-Stage DNA Operation, MuStDO) that includes a liver-stage DNA vaccine candidate encoding the CS protein of P. falciparum. This DNA vaccine tested in Phase I showed no serious adverse events and no detectable DNA autoantibodies after a one-year follow-up but the vaccination failed to induce antigen-specific antibodies.

A multiple-antigen version of the DNA vaccine, MuStDO 5, encodes five different liver-stage antigens including CS and the additional antigens: liver stage antigens 1 and 3 (LSA 1 and 3), exported protein 1 (EXP1), all having shown to be protective in mouse models, and sporozoite surface protein 2 (SSP2, previously tested in animals as part of a DNA vaccine mix and also known as thrombospondin-related adhesive protein, TRAP). MuStDo 5 is manufactured as a combination of five separate plasmids. The DNA vaccine administered with GM-CSF DNA as adjuvant was safe and well tolerated in mice and rabbits. The LSA 3 protein has been demonstrated to induce protective immunity against P. falciparum infection in the chimpanzee model. Various formulations of this antigen (peptides, lipopeptides, and DNA vaccine) are currently targeted for clinical development.

Several groups are using a prime-boost approach by priming with a DNA vaccine and boosting with either recombinant antigen or viral vectors, shown to be more immunogenic than either vaccine alone.

Another vaccine development approach targeting the pre-erythrocytlc parasite is focused on the intracellular liver stage parasite. Several known antigens expressed by sporozoites or merozoites can also be expressed by liver stage parasites. Various studies conducted in endemic areas have linked liver stage antigen 1(LSA-1) with protective immunity. B-cell epitopes in LSA-1 and LSA-3 have been associated with protective immune responses in these studies.

Additional antigens that have been targeted for vaccine development because of identification of epitopes associated with protective immunity include the sporozoite and liver stage antigen (SALSA) ,sporozoite threonine and aspargine rich protein (STARP - also expressed in sporozoites) , and the glutamate-rich protein (GLURP) . The progress in the field of peptide synthesis now permits the synthesis of long chains of synthetic peptides (LSP) which allows for the inclusion of multiple protective epitopes. This long synthetic peptide approach to malaria vaccine design is being pursued by several collaborative groups in Europe and Africa.

Asexual blood-stage vaccine candidates. The most advanced asexual blood stage vaccine is merozoite surface protein 1 (MSP1). MSP1 forms part of a complex that is thought to be involved in red blood-cell invasion and antibodies to MSP1 have been shown to block parasite invasion of red blood cells in-vitro. MSP1 contains the presence of two connected cysteine-containing epidermal growth factor (EGF)-like modules required to maintain the conformation-dependent epitopes needed to generate protective antibodies. The EGF-like domains is conserved across all species of Plasmodium, thus this region is a prime focus of current MSP1 vaccine candidates. Several groups work on MSP1 either as the entire molecule, the 42kDa C-terminal moiety, the further-processed 19kDa fragment, or as part of a hybrid molecule. BaculovIrus, E. coli, and yeast (Saccharomyces or Pichia) are the expression systems used for recombinant antigen production. Recombinant MSP1 (42kDa or 19kDa), produced in each of these different systems, has been shown to protect both mice and monkeys against lethal parasite challenge. Several DNA vaccine constructs are also being tested. Most of these vaccines are in pre-clinical stage of optimization prlor GMP manufacturing for Phase I/II trials.

A number of additional merozoite surface protein (MSP) antigens are under development as vaccine candidates (MSP2, 3, 4, 5, 8 and 9). These related molecules contain one or more of the hallmark EGF-like domains present in MSP1. MSP5 is of particular interest because it lacks the sequence variation between different isolates of P. falciparum from different geographical locations (typically seen with most of the merozoite surface proteins), which may simplify vaccine formulation.

Currently most advanced along the vaccine development pathway of blood-stage malaria vaccine candidates is the 'Combination B' vaccine candidate. This vaccine combines MSP-1 and MSP-2 with Plasmodium falciparum ring-infected erythrocyte(RESA). Recently, phase I/IIb trials of this vaccine In Papua New Guinea children aged 5-9 showed a 62 % reduction in parasite density In participants who were not pre-treated with sulfadoxine-pyrimethamine before vaccination . This trial highlighted the issue of clearing parasitemia before vaccination with treatment as the efficacy of the vaccine was only significant in the group who were not pre-treated. It also demonstrated possible vaccine-induced selective pressure on the MSP-2 component of the vaccine underlining the importance of development strategies that focus incorporating all significant genotype or highly conserved antigens in vaccine design.

Two other promising P. falciparum asexual blood-stage candidate antigens are the apical membrane antigen-1 (AMA-1) and erythrocyte binding antigen-175 (EBA-175). There are currently 4-5 different laboratories developing these antigens as vaccine candidates expressed either in E. coli, Pichia pastoris

or as a DNA prime-boost vaccine.

Transmission-blocking vaccine candidates. The leading candidate vaccines contain the P. falciparum surface protein antigens Pfs 25 and Pfs 28 or the P. vivax homologues referred to as Pvs25 and 28. Currently, transmission-blocking antigens Pvs28, Pvs28, and Pfs25 are being developed at the NIH (USA) as recombinant yeast-secreted proteins (S. cerevisiae). Initial human Phase I safety, immunogenicity and in vitro efficacy trials have been done for Pfs25 and might be following soon for Pvs25. Other sexual stage-specific antigens that are being developed as transmission-blocking vaccines are Pfs48/45 and Pfs230.

Anti-toxic candidate vaccine - the anti-GPI vaccine Anti-disease vaccines are also being developed that involve direct immunization against parasite toxins that are identified as the cause of disease pathology. The identification of these toxins must be followed by characterization of the immune response to the toxin, focusing on it's subsequent neutralization by antibodies and most importantly, prevention of disease pathology, in order to be considered as a potential anti-disease malaria vaccine candidate.

The glycosylphosphatidylinositol (GPI) anchor, which tethers several of the parasite antigens to the membrane, has been shown to be highly toxic in mouse models, and is currently being developed as a carbohydrate anti-toxic vaccine. A study testing synthetic GPI in rodent models of malaria appeared to show that the candidate anti-toxic vaccine was immunogenic and protected the rodent model from significant malaria pathologies and mortalities . Following this initial proof-of-principle in an animal model, further development of the malaria toxin neutralization as a vaccine strategy continues.

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State of the art of new vaccines: research & development: 1,2,3,4,5,6,7,8,9,10,11 | Next page

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DOCUMENT II

Portfolio of candidate malaria vaccines currently in development February 2004

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to Zarifuh Reed (reedz@who.int)

Activities completed or ongoing are marked with a bold X Support for activities indicated with colour and without an X have been secured The various stages are considered as initiated at the steps indicated below:

Preclinical development stage: serious process development has been initiated

Phase 1a: first volunteer recruited for phase 1 in industrialized country
Phase 2a: first volunteer recruited for challenge study in industrialized country

Phase 1b: first volunteer recruited for phase 1 in disease endemic country

Phase 2b: first volunteer recruited for phase 2 in disease endemic country

Pivotal: first volunteer recruited for clinical study leading to licensure

Pre-erythro	ocytic va	accines					
	Research	Preclinical development	Phase 1a	Phase 2a	Phase 1b	Phase 2b	Pivotal
CSP		,		1	1		
HBSAg-CSP chimeric mixed VLP (RTS,S)/AS02A (GSK)	X	X	X	X			
HBSAg-CSP chimeric mixed VLP (RTS,S)/AS02A and (RTS,S)/AS01B (WRAIR/GSK)							
HBCAg-CSP VLP (Malarivax Apovia)	20,110	a basilian sa	in the Prince and				
Modified Vaccinia Ankara (MVA) CSP tested in combination with RTS,S/AS02 (Oxford-GSK)	X			Ţ			
Recombinant adenovirus CSP (NYU)							
Recombinant adenovirus CSP (WRAIR/Crucell Holland)		Ţ.					
Recombinant influenza CSP (NYU)		·	# × <u> </u>	<u> </u>			ļ
Recombinant vaccinia CSP (NYU)							
Recombinant Sindbis virus CSP (NYU)				<u>L</u>			
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re-erythrocytic vaccines – CSP (continued)	Research	Proclinical development	Phase a	Phase 2a	Phase 1b	Phase 2b	Pivotal
ecombinant Yellow Fever virus CSP	X						
IVIO		X	X	W.E.			
ong synthetic CSP peptide		^					1
ausanne)		N,	X			1	1
SP DNA immunization	X	X	· ^			1	1
NMRC)							1
			X	-		+	1
ong Vivax CSP C-terminus 72 AA Synthetic	X	X .	L			↓	-
eptide (MVDC) ong Vivax CSP N-terminus 77AA Synthetic peptide	X	X	X _				
MVDC)	X	X	X				
ong Vivax CSP repeat 48 AA Synthetic peptide							
MVDC)		<u> </u>					
Other antigens	X					1	
ong synthetic LSA-3 peptide	1			1			
Pasteur Institute)		Part Heathern Control	4.7				
Lactis expressed recombinant LSA-3 protein +	X	X			·		1
ipopeptides (Institut Pasteur)			-				
E. coli recombinant LSA-3 (WRAIR)		-					
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E. coli recombinant LSA-1(LSA-NRC)	हेड संस्था के सिसंहर के स						
(WRAIR)	W. C. W.						}
Adenovirus vectored LSA-1(LSA-NRC)	70			,			-
(WRAIR/Crucell Holland)							
Modified Vaccinia Ankara (MVA) CSP + LSA-1							
epitope (Oxford)							
Fowl Pox 9 CSP + LSA-1 epitope					6.		
(Oxford) Fowl Pox 9 CSP + LSA-1 epitope/ Modified	X	X	P			1	
Fowl Pox 9 CSP + LSA-1 epitoper Modified Vaccinia Ankara (MVA) CSP + LSA-1 epitope	M -		7		Ĭ		1
	Щ		_ 3		S		
(Oxford) DNA MVA prime-boost Multi-epitope(ME) string +							
					L		
TRAP (Oxford)							
Fowl Pox 9 MVA prime-boost ME string + TRAP							
(Oxford)							
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	Preclinical	Phase	Phase	Phase	Phase	Pivotul
	development.	-1a	2a	16	2b	+
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age vace	Perlinien	Phase	Phase	Phase	Phase	Pivotal
Research		1	2a	1b	2b	
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Blood Stage vaccines - MSP-1 (continued)	Research	Preclinical development	Phase la	Phase 2s	Phase 1b	Phase 2b	Pivotal
P pastoris expressed recombinant protein MSP-1 42Kd 3D7 (MVDU)							
P pastoris expressed recombinant protein MSP-1 42Kd FVO (MVDU)							
P pastoris recombinant AMA-1 and MSP-1 chimera (SMMHS)							
E. coli recombinant MSP1-42-kDa-EBA-175 chimera (WRAIR)	X	X					
E. coli expressed recombinant MSP1-19 and EBA- 175 F1 domain protein (ICGEB)							
D. melamogaster expressed MSP1 19 kD protein (Hawaii Biotech, Inc.)	=		Sandina				
D. melamogaster expressed MSP1 42 kD protein (Hawaii Biotech, Inc.)							
BCG vectored MSP-1 (AECOM)							<u></u>
Salmonella-vectored MSP-1 (U. Maryland)							
Other MSPs							
Long synthetic peptide MSP-2 (Lausanne)	X	X					
E. coli expressed recombinant protein MSP-2 3D7+ (FC27) (La Trobe)	X						
Long synthetic peptide MSP-3 (Pasteur Institute)	X		Y		X	-	
E. coli expressed recombinant MSP-4 protein (Monash)	7					i	
E. coli expressed recombinant MSP-5 protein (Monash)							
MSP-3-GLURP hybrid vaccine (SSI)	X				X		
AMA-1		*** **********************************					
E. coli recombinant protein AMA-1 (Australia)							
E. coli expressed recombinant protein AMA-1 3D7 (MVDU)	200000000000000000000000000000000000000	tin Samuel Territor Fills of The	A Ser hims and her him				

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Blood Stage vaccines - AMA-1 (continued)	Research	Preclinical development	Phase la	Phase 2a	Phase 1b	Phase 2b	Pivota
E. coli expressed recombinant protein AMA-1 FVO (MVDU)							
P pastoris expressed recombinant AMA-1 protein (BPRC)		2017					
E. coli recombinant protein AMA-1 FVO (FMP10)(WRAIR)		10 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1					
E. coli recombinant protein AMA-1 3D7 (FMP2.1)(WRAIR)							
EBA-175 and DBP							
E. coli expressed recombinant Region II Duffy Binding Protein protein (ICGEB)			=				
E. coli expressed recombinant EBA-175 F1 domain protein (ICGEB)					X		
P pastoris recombinant protein EBA-175 (F1 +F2) (EntreMed/SAIC)		v! 24	:				
Other proteins							
Long synthetic GLURP peptide (SSI)	X						
MAEBL (Notre Dame University) P. Pastoris expressed erythrocyte binding proteins	~						
EBP2/BAEBL (Entremed)							
E. coli expressed recombinant RAP-2 protein (QIMR)	X						
E. coli, P. pastoris, baculovirus expressed PfEMP1, different domains (various EU groups)		X					
P. falciparum synthetic GPI toxin	8		1				
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Transmiss	ion blocking	g vaccines					
	Research	Preclinical development	Phase 1a	Phase 2s	Phase 1b	Phase 2h	Pivota
Saccharomyces recombinant protein PvS25 (MVDU)							
P Pastoris recombinant protein PfS25 (MVDU)							
Recombinant protein PfS48 (MVDU)							
Recombinant protein PfS48 (Nijmegen)						<u> </u>	
DNA immunization PfS25 (JHU)							
DNA immunization PvS25 (JHU)		*					
DNA immunization PvS28 (JHU)							
Recombinant Pfs230 (Loyola University)		,					

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	Research	Preclinical development	Phase la	Phase 2a	Phase 1b	Phase 2b	Pivotal
Multi-epitope recombinant protein CSP, MSP-1, MSP-	X					ļ	
2, LSA-1, AMA-1, RAP-1, EBA-175 (FALVAC CDC)		1		Annual Control		ļ	├ ~~
MVA prime-boost Fowl Pox 9 LSA3/D260; STARP;		na managanga ku singan pengangan beragangan					1
EXP1, Pfs16, TRAP, LSA-1 (Oxford)	X						
DNA in co-block polymer +/- viral boost CSP, SSP2,					Y		
LSA-1, AMA-1, MSP-1 (NMRC)				ļ			
MVA CSP, SSP2, LSA-1, AMA-1, MSP-1	500				'		1
(NMRC)						<u> </u>	
Recombinant Salmonella-vectored vaccine CSP,							
SSP2, LSA-1, MSP-1, individually as well as in				ļ į			İ
combination (U Maryland)		k					
Recombinant Shigella-vectored vaccine CSP, SSP2,						Ì	1
LSA-1, MSP-1, individually as well as in		and a					
combination (U Maryland)							
Multivalent antigen expression/vaccine for malaria						1	
(Lifesensors, Inc.)		E.				ļ	
Recombinant Adenovirus CSP, SSP2, LSA-1, AMA-	X	X					
1, MSP-1 (NMRC/Genvec)							
Recombinant FMP-1 plus RTS,S, MSP-1 3D7 +	Art the		,	1			
CSP (WRAIR)							
Mimetopes delivered on Virosome CSP, MSP-1,	X	X					
AMA-1 (Pevion)						<u> </u>	L

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